

Decomposition of Overlapping Protein Complexes

A Graph Theoretical Method for Analyzing Static and Dynamic Protein Associations

Teresa Przytycka

NIH / NLM / NCBI

joint work with

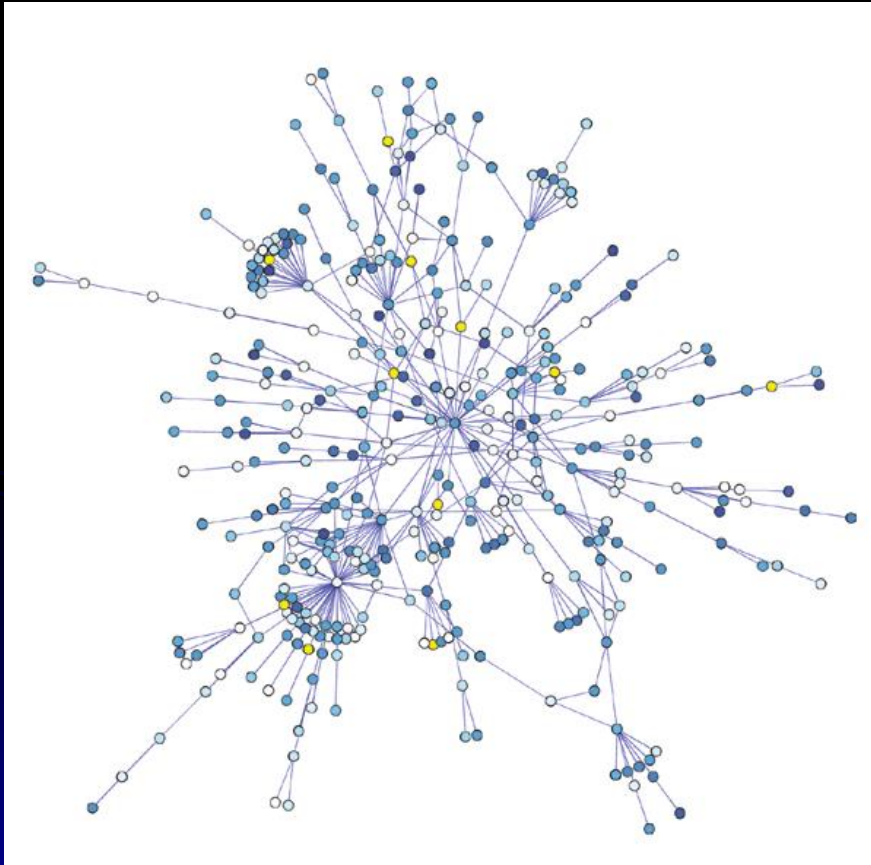
Elena Zotenko

Katia S. Guimaraes

Raja Jothi



Investigating protein-protein interaction networks



- **Scale free vertex degree distribution**

(Barabasi & Albert 1999; ...)

- **Modular organization**
- **Recognizing functional modules**

(Spirin & Mirny 2003, Rives & Galitski 2003, Bader *et al.* 2003, Bu *et al.* 2003,...)

- **Representing variants of protein complexes**

(Gagnier *et al.*, 2004)

Image by Gary Bader (Memorial Sloan-Kettering Cancer Center).

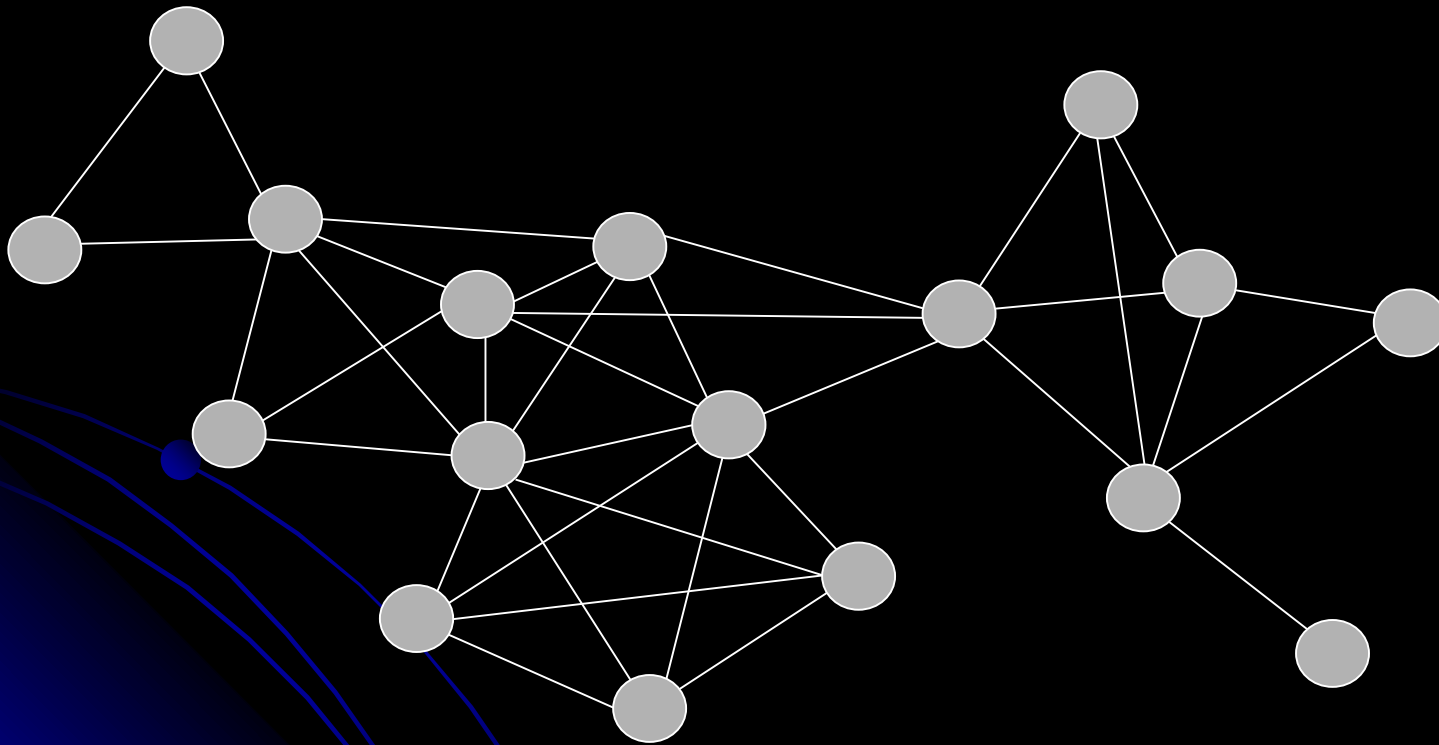
RECOMB Systems Biology, December, 2005

Functional Modules and Functional Groups

- **Functional Module:** Group of genes or their products in a metabolic or signaling pathway, which are related by one or more genetic or cellular interactions and whose members have more relations among themselves than with members of other modules (Tornow *et al.* 2003)
- **Functional Group:** protein complex (alternatively a group of pairwise interacting proteins) or a set of alternative variants of such a complex.
- Functional group is **part of functional module**

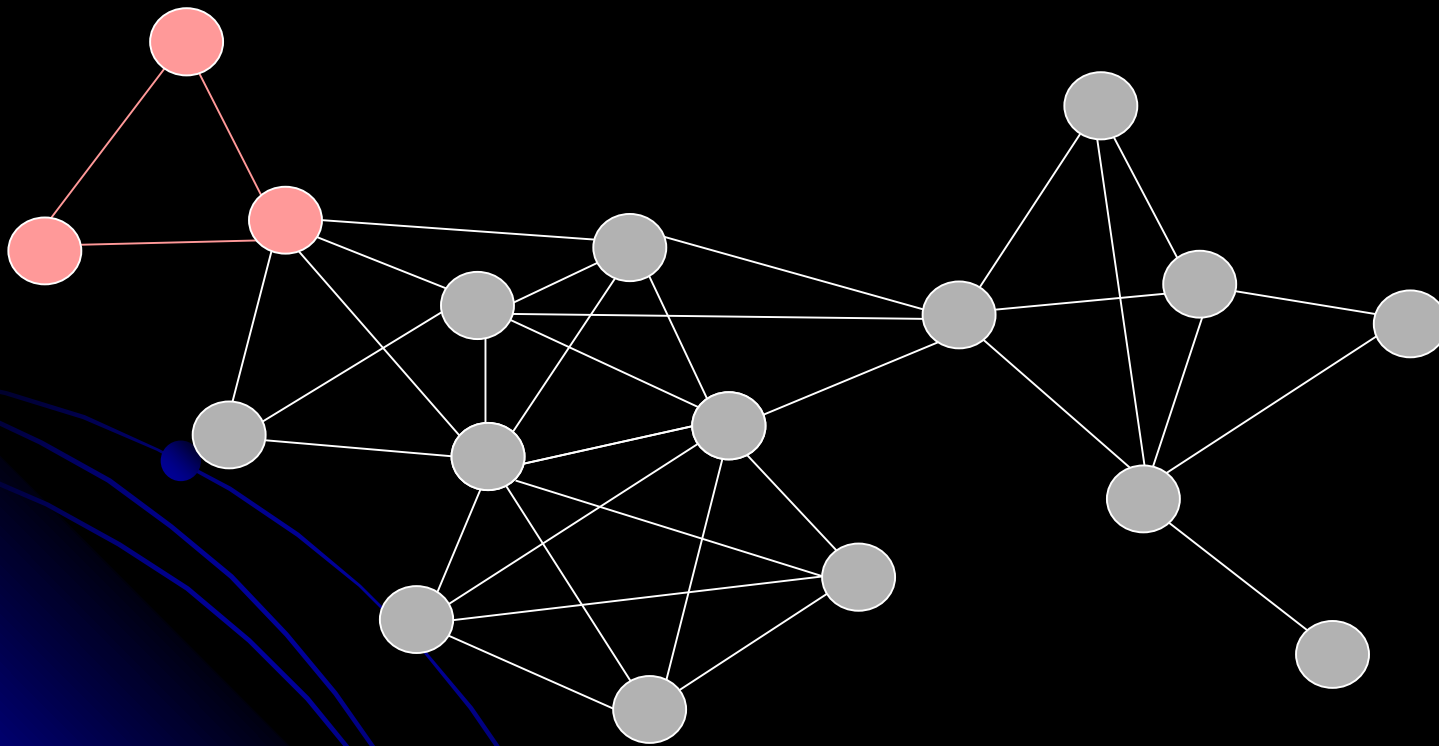
Overlaps between Functional Groups

For an illustration functional groups = maximal cliques



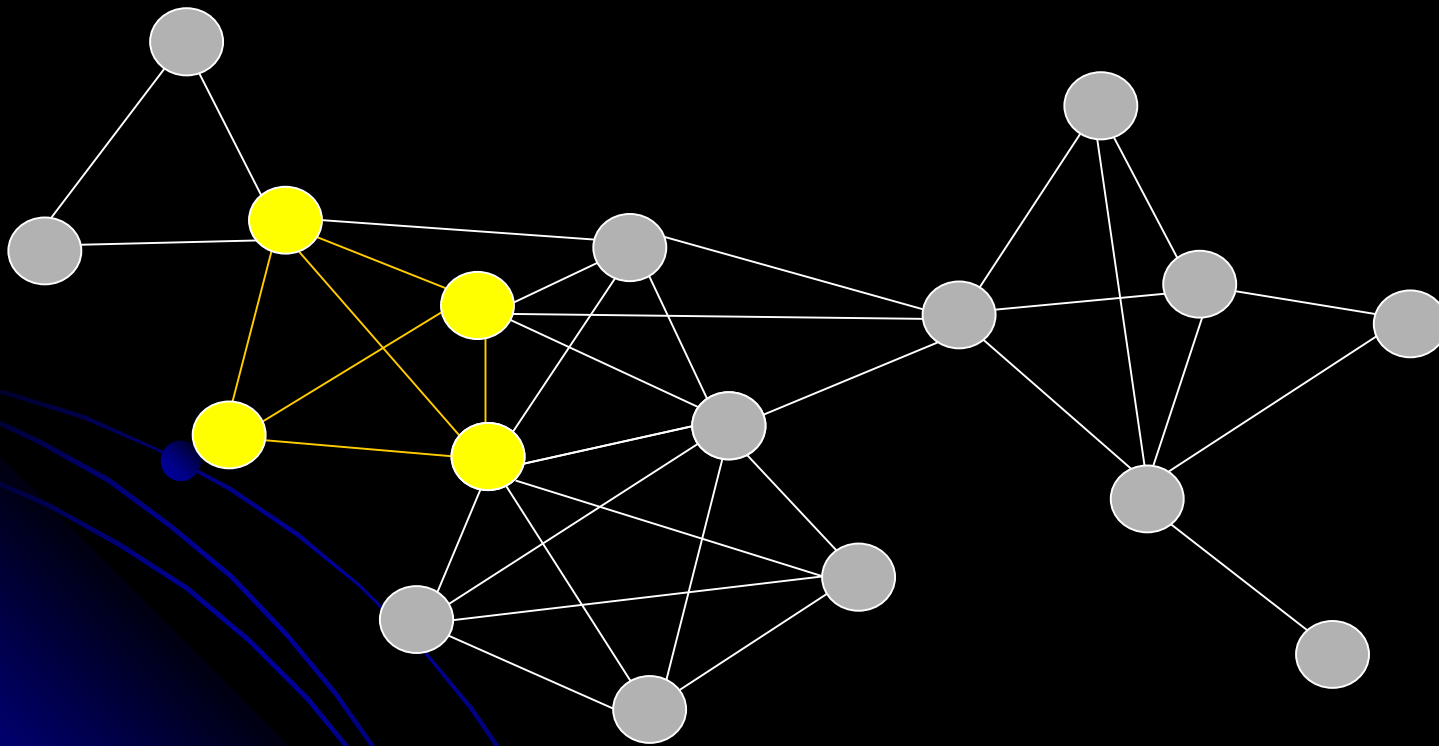
Overlaps between Functional Groups

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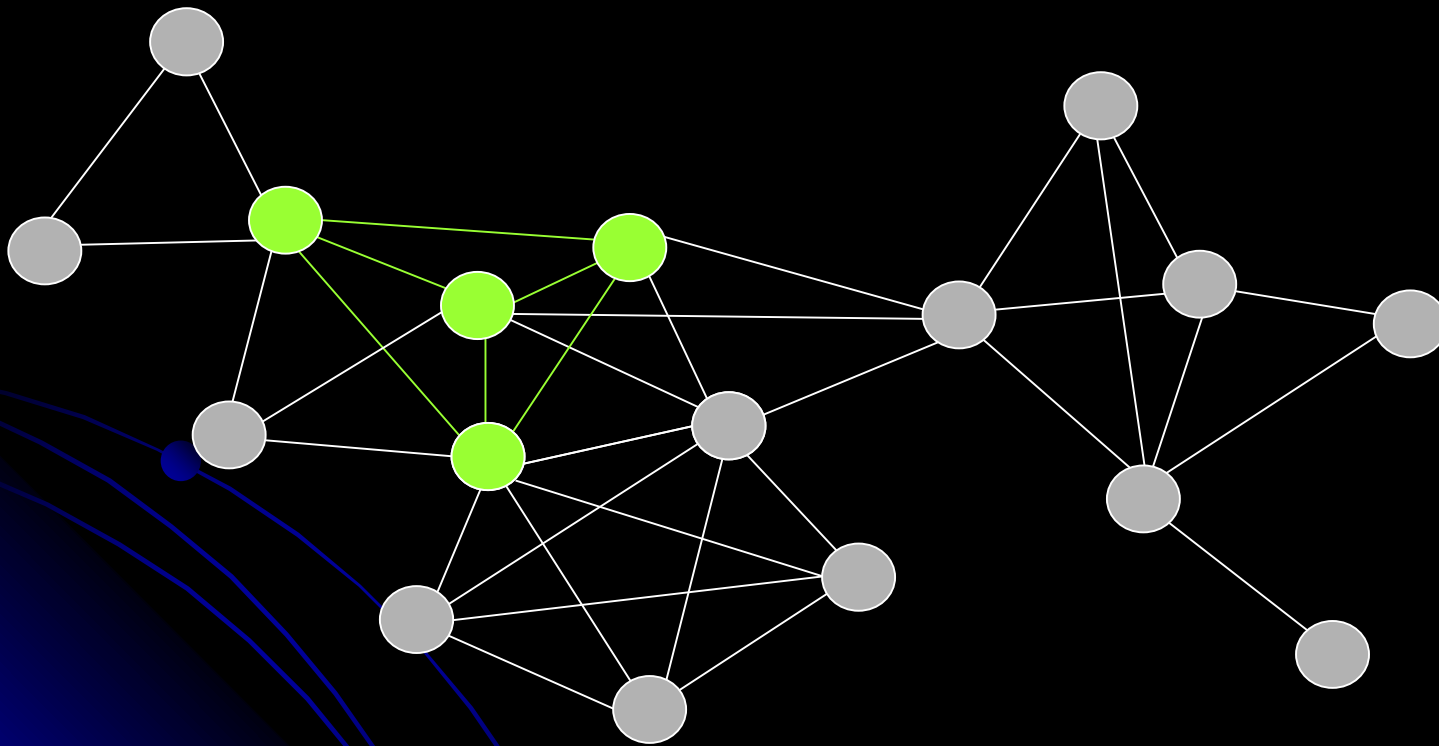
Overlaps between Functional Groups

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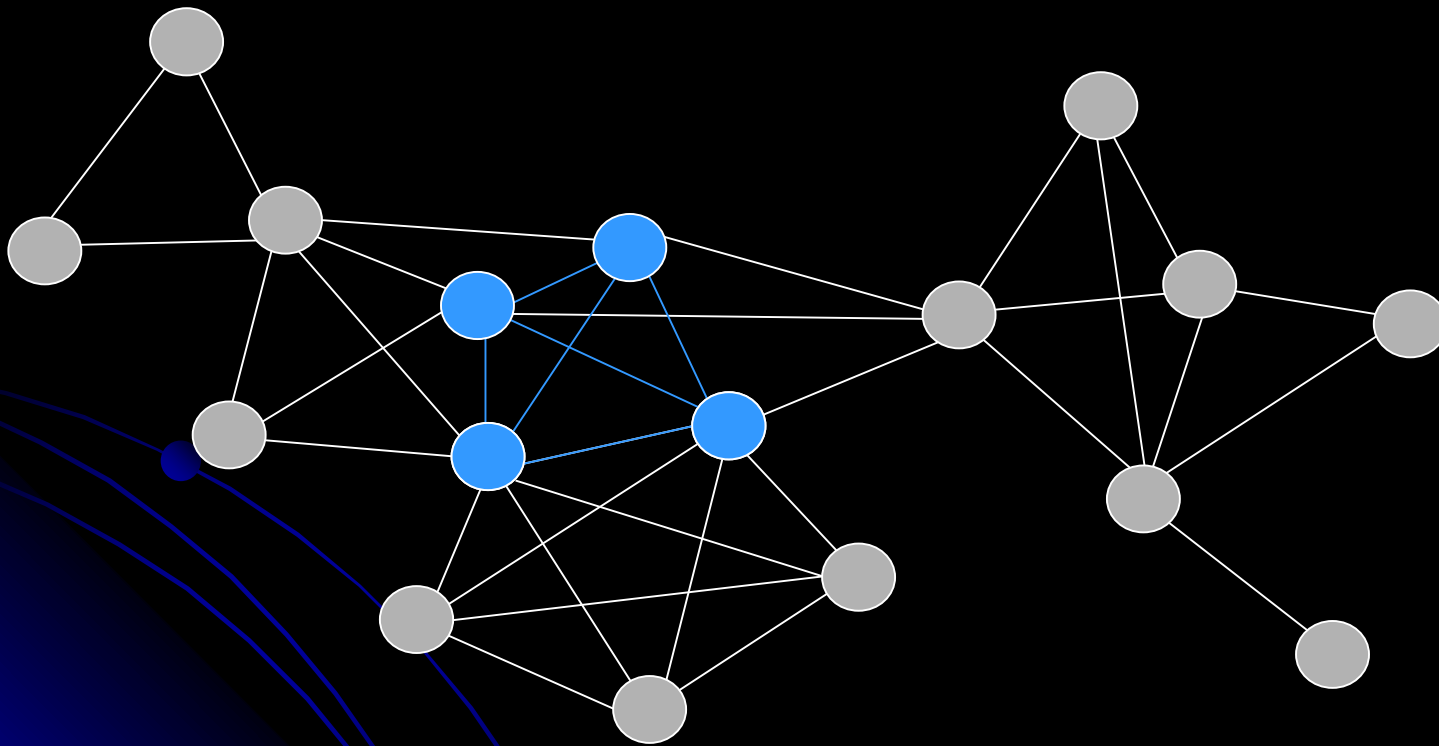
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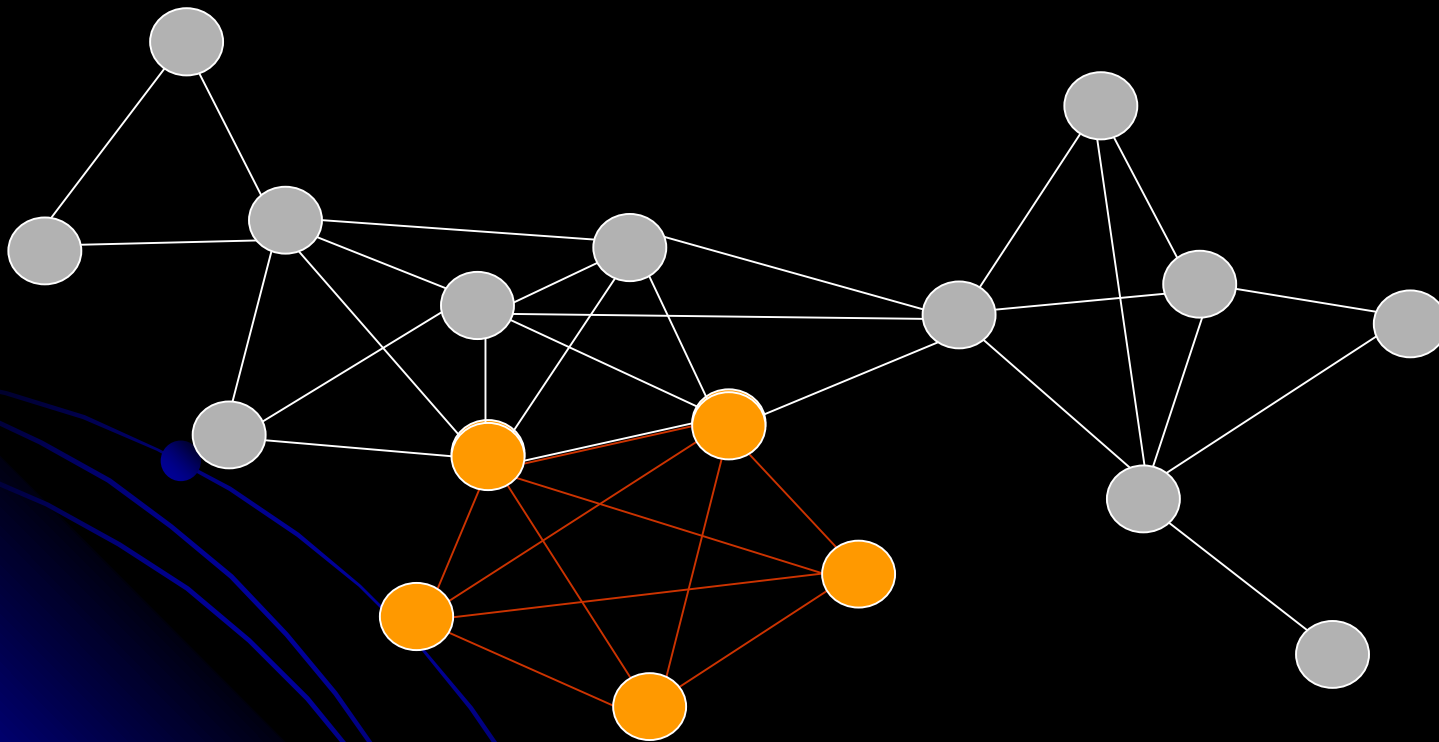
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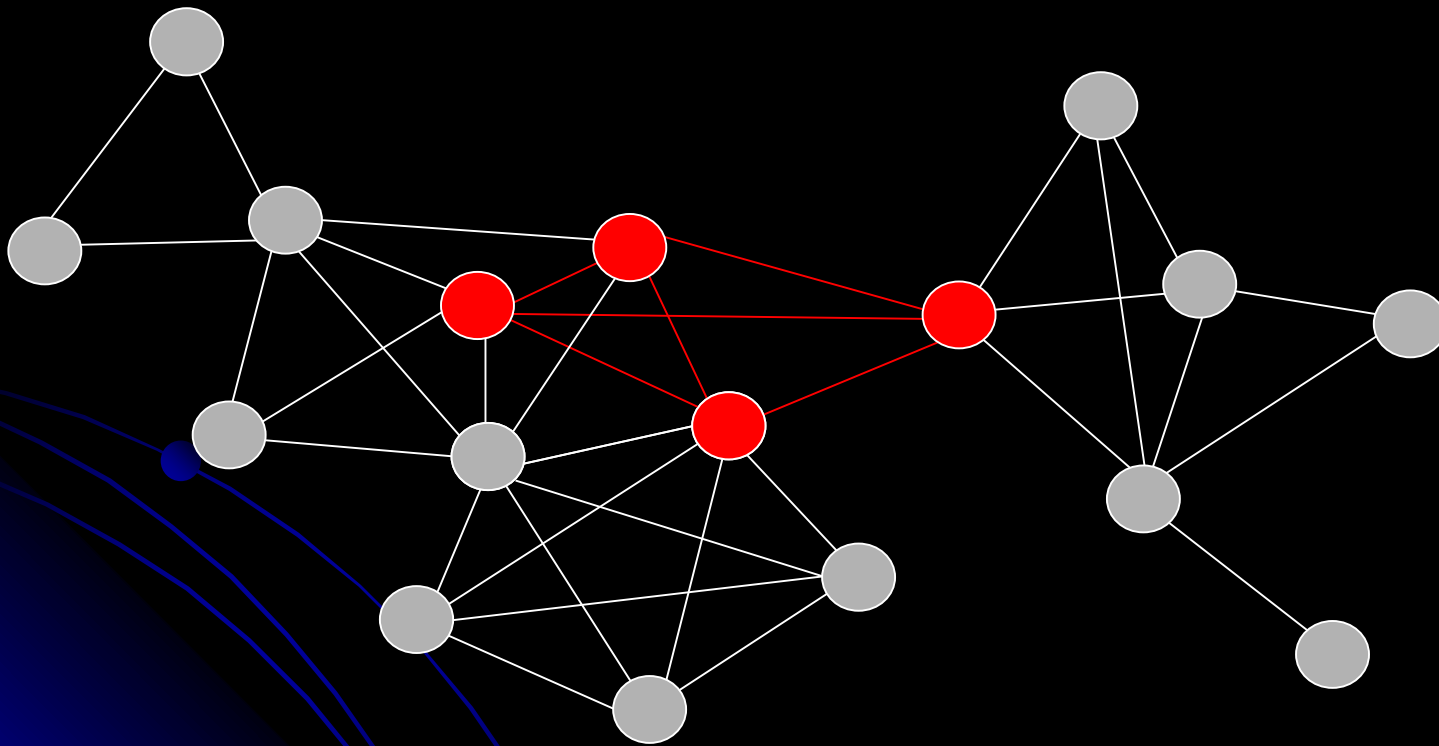
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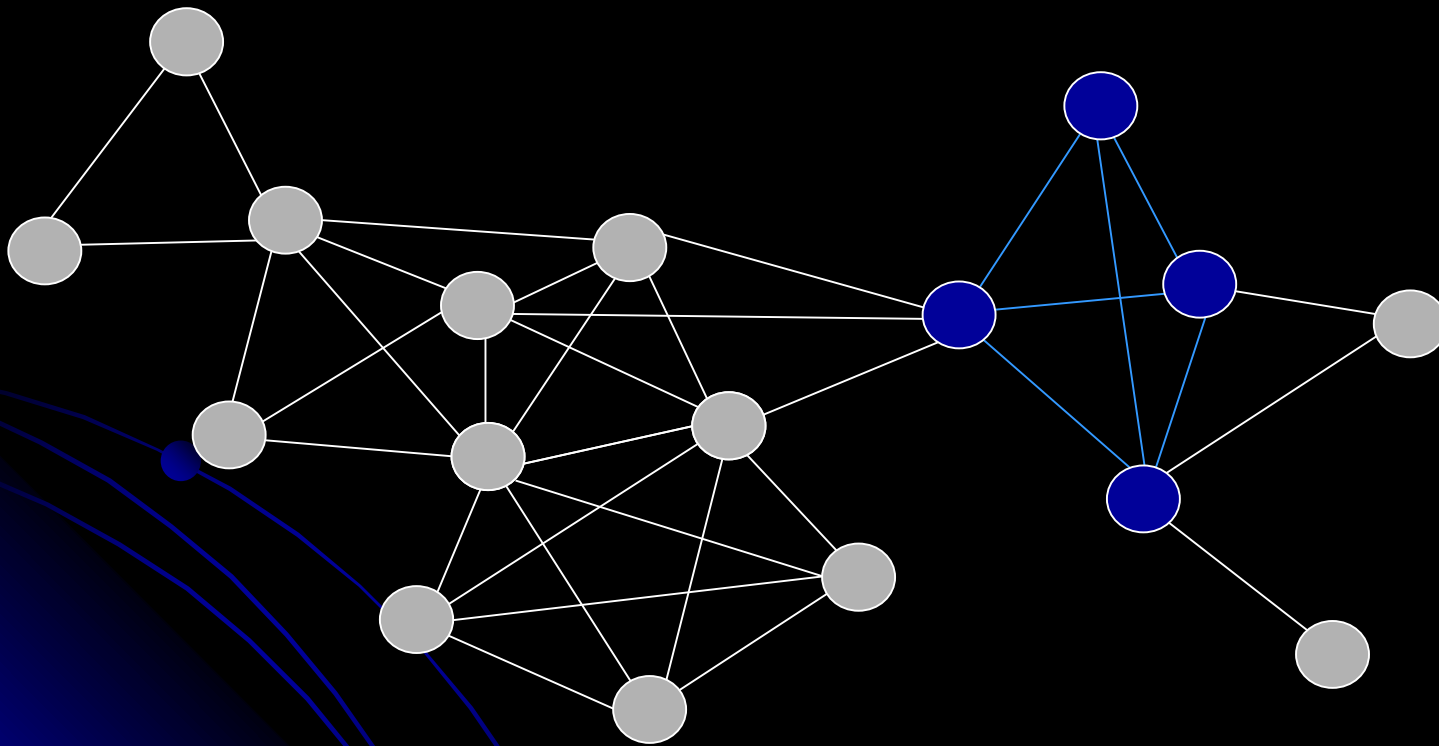
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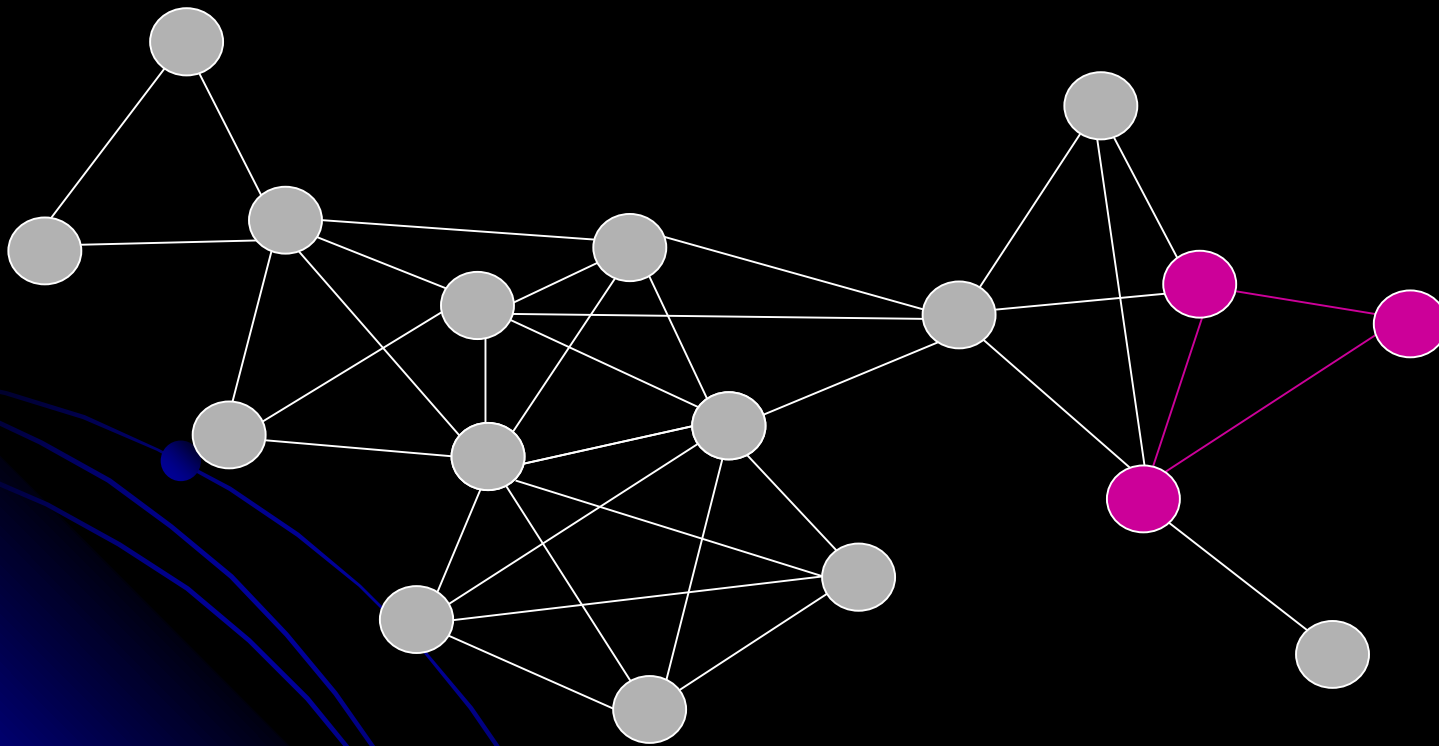
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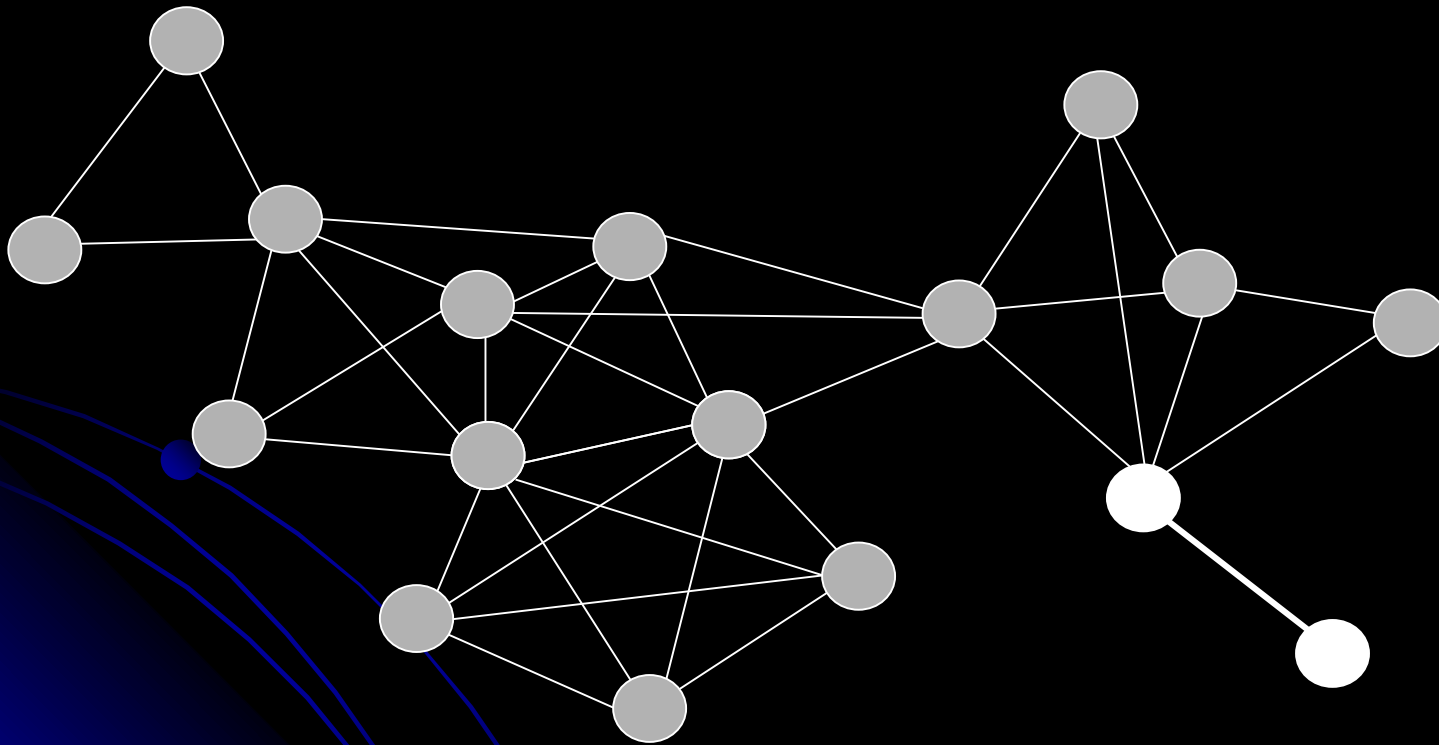
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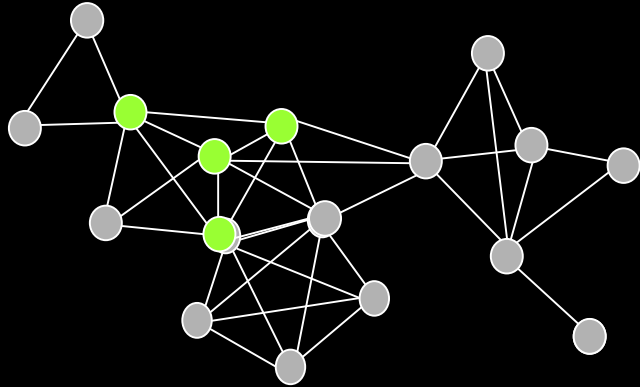


Overlaps between Functional Groups

For an illustration functional groups = maximal cliques



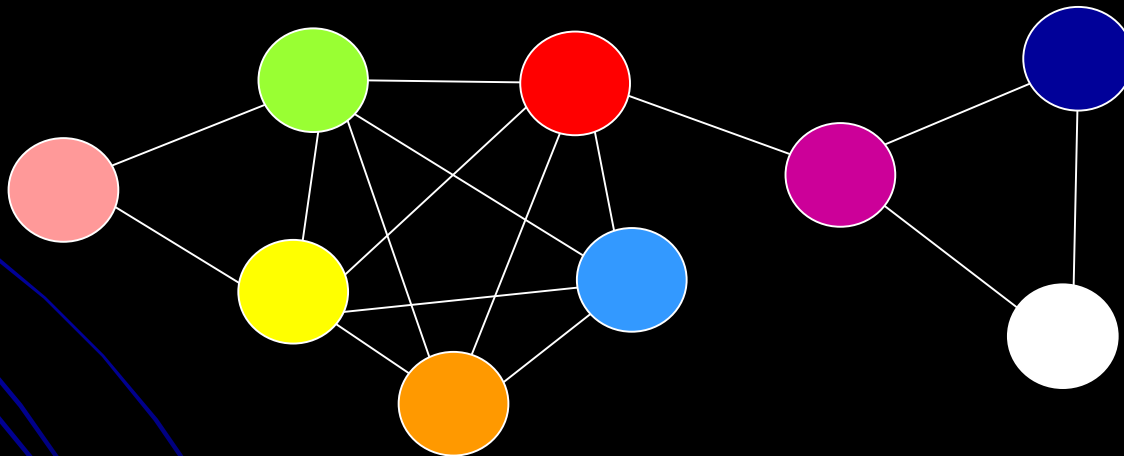
First line of attack



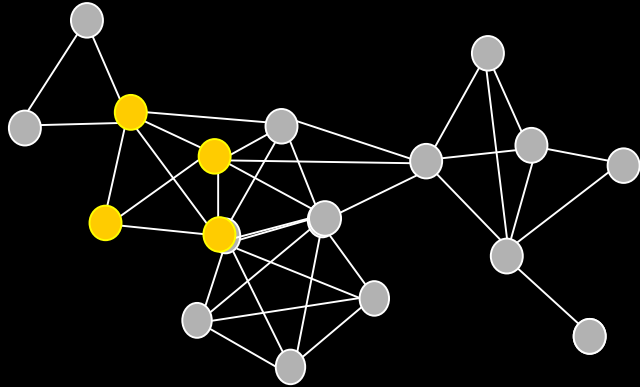
Overlap graph:

Nodes= functional groups

Edges= overlaps between them



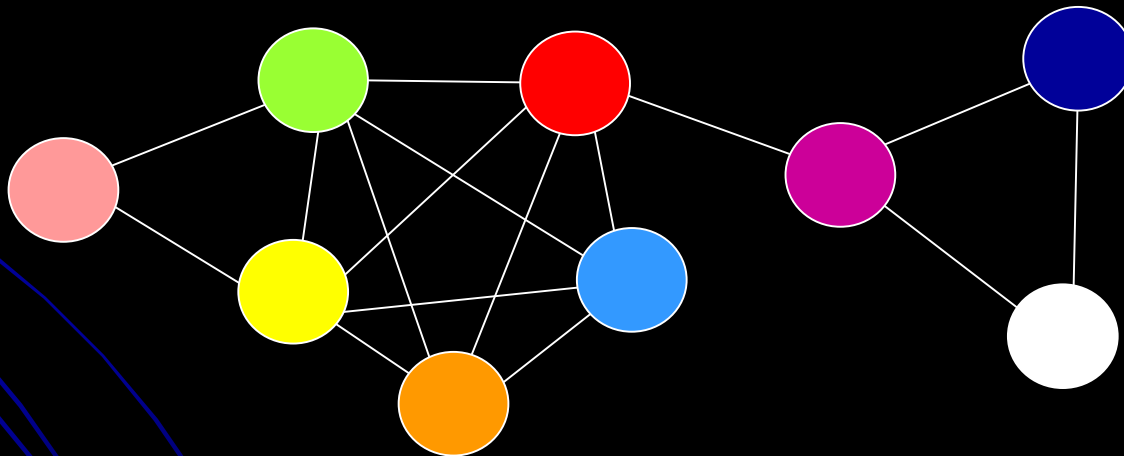
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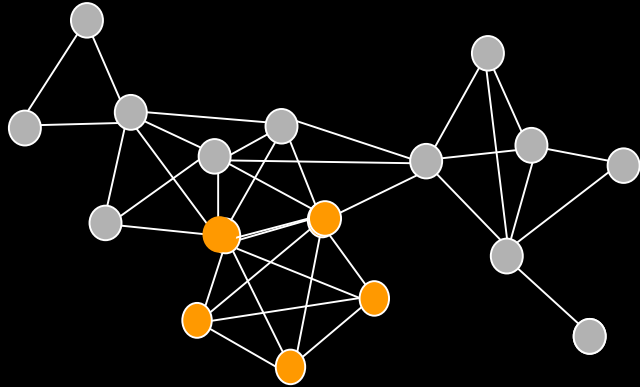
Overlap graph:

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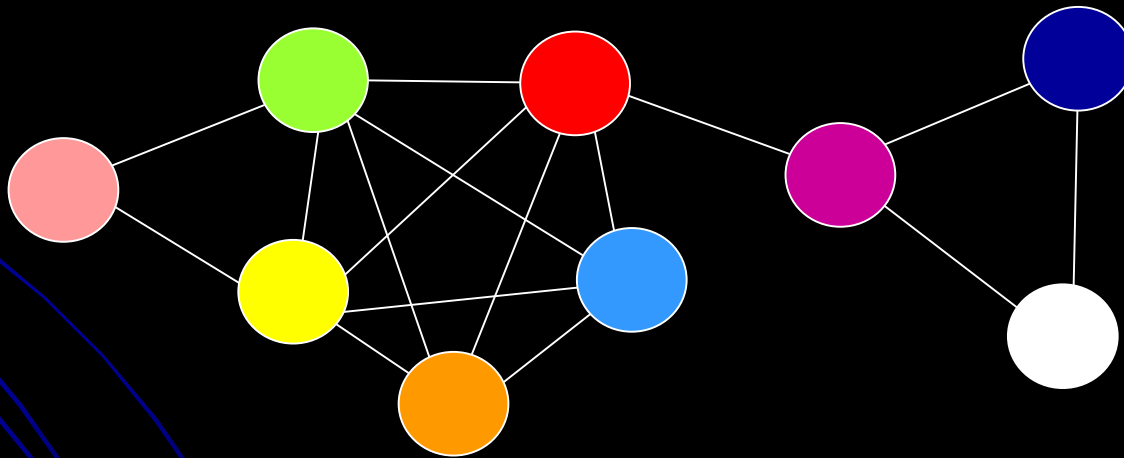
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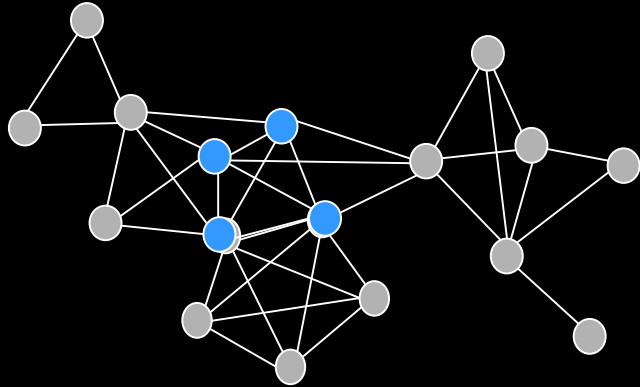
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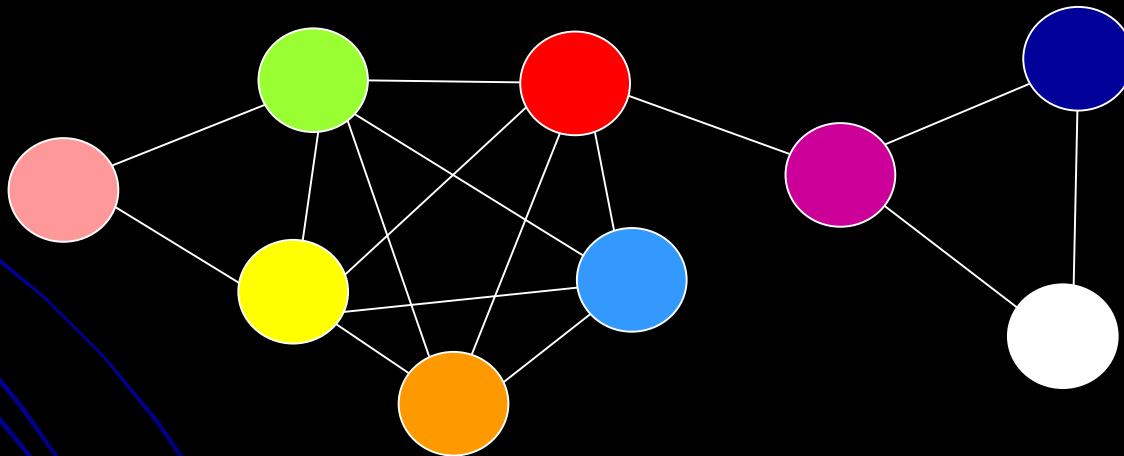
First line of attack



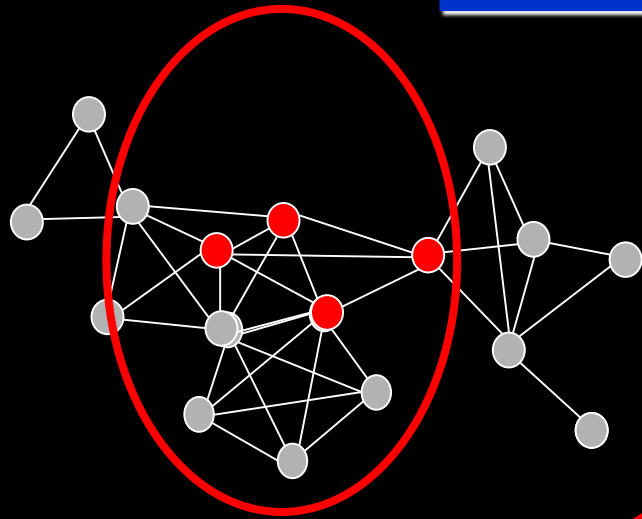
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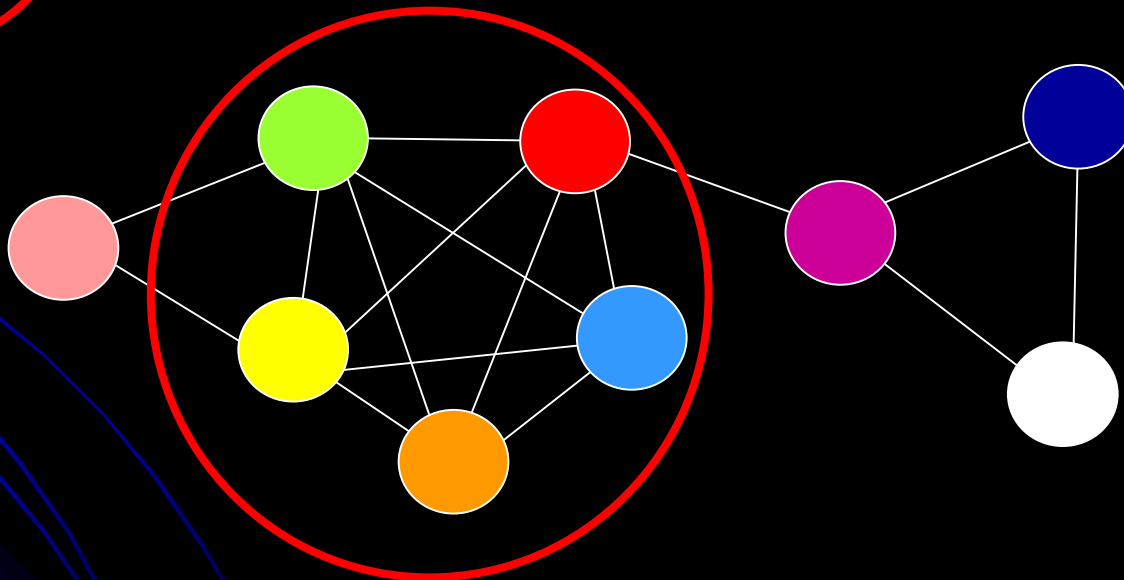
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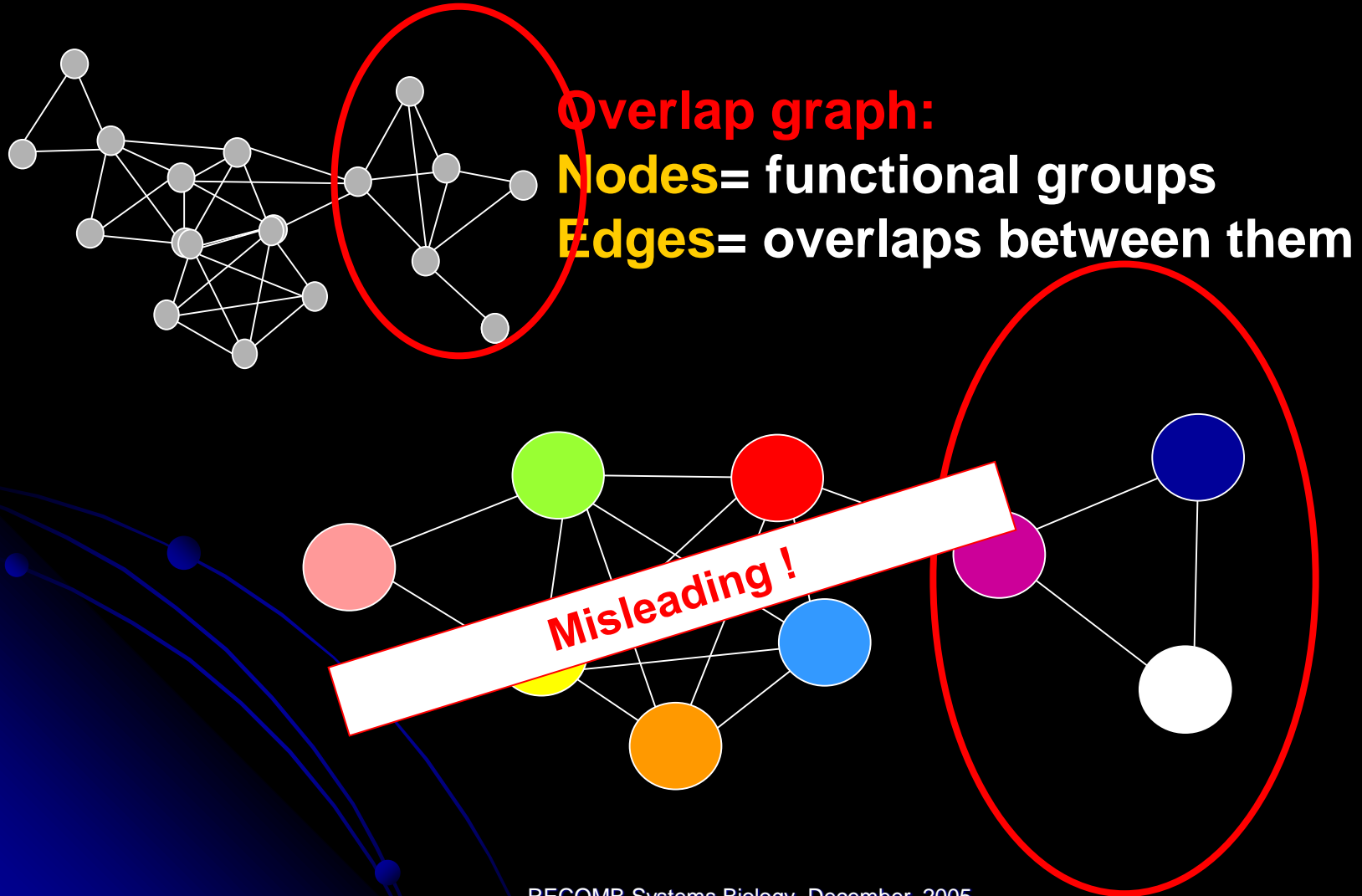
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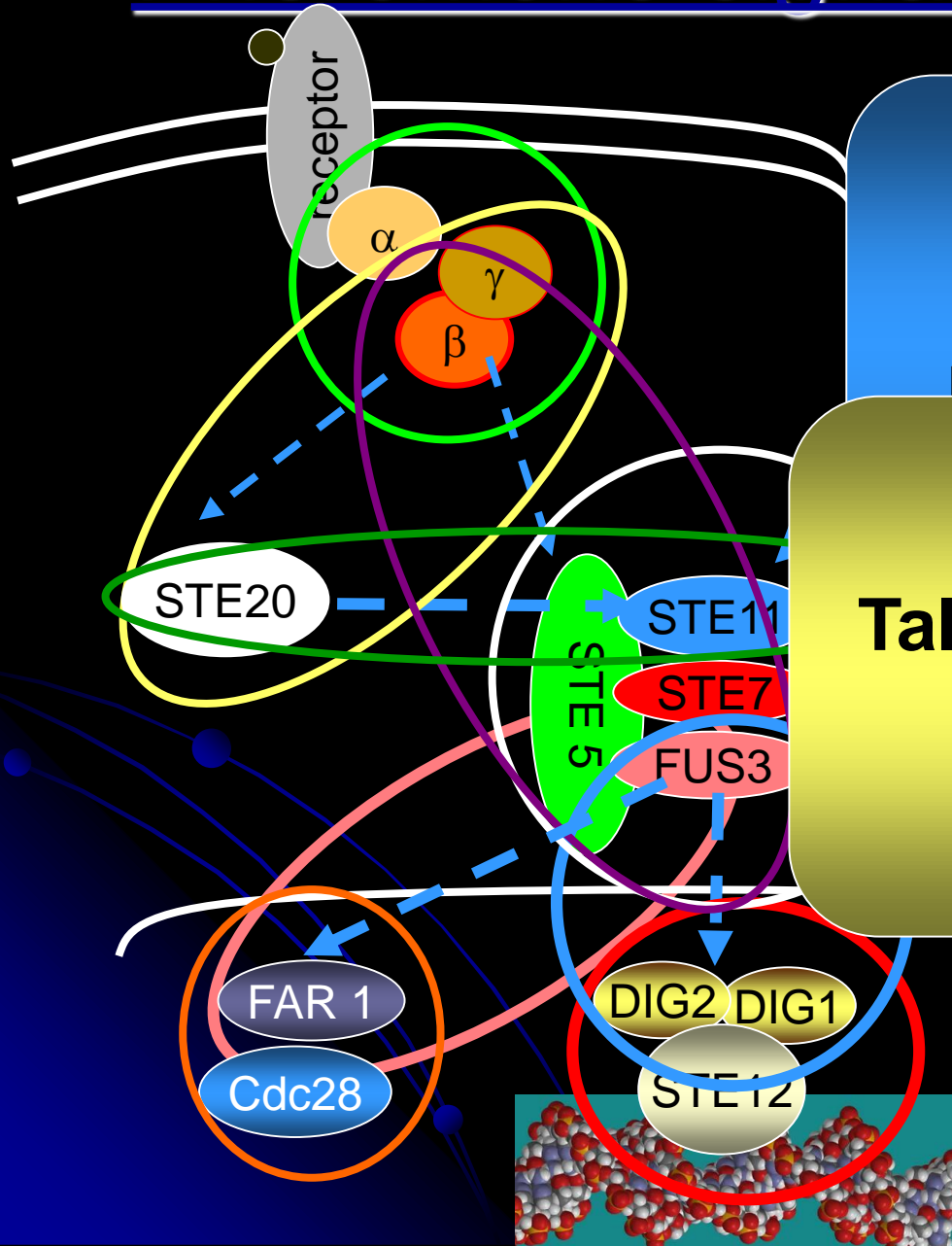
First line of attack



Our contribution

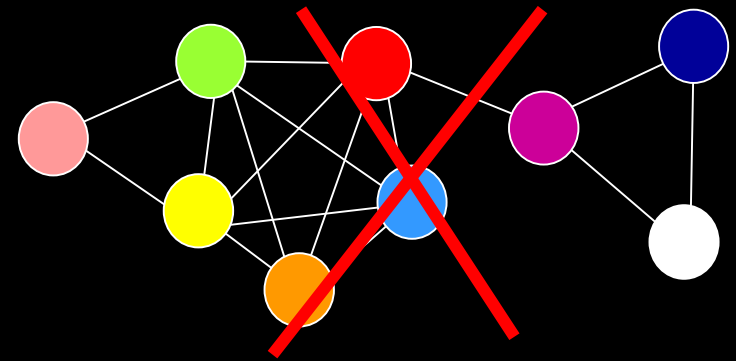
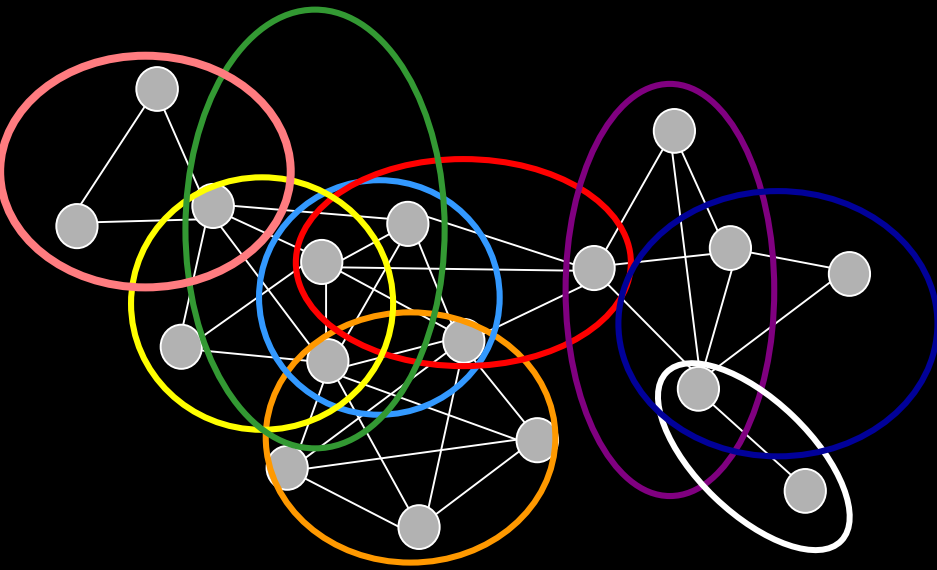
- Approach that elucidates the structure of the overlaps
- Application: If the functional module represents dynamically changing protein associations, the method can suggest temporal relations between these associations

Pheromone signaling pathway



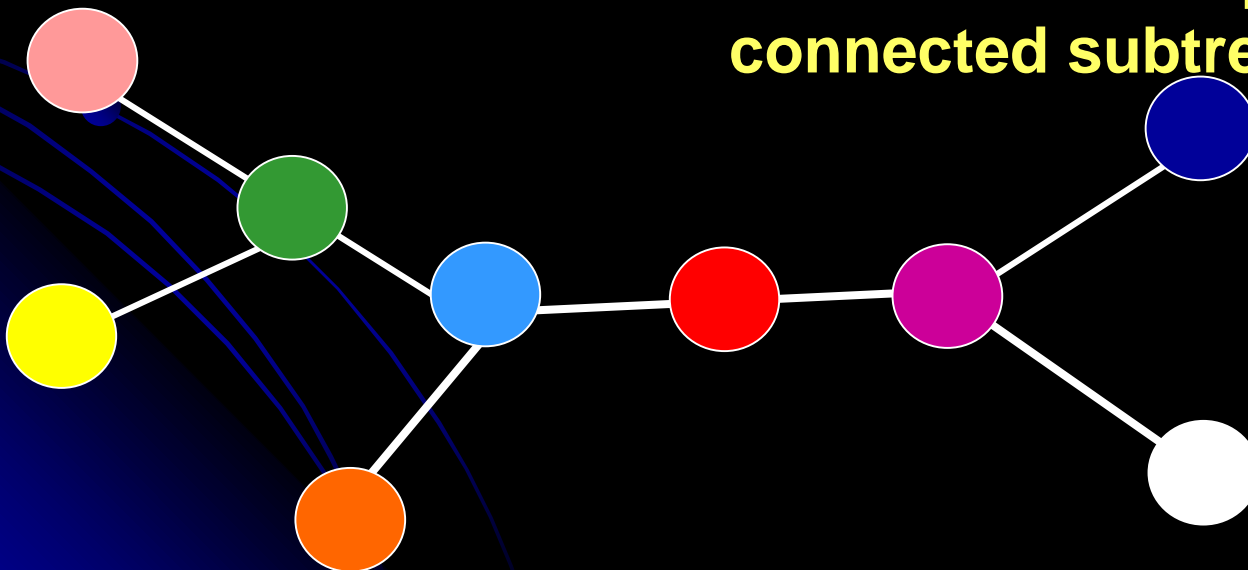
The information about overlaps be used to propose a partial order of

Talking partial order =
talking trees

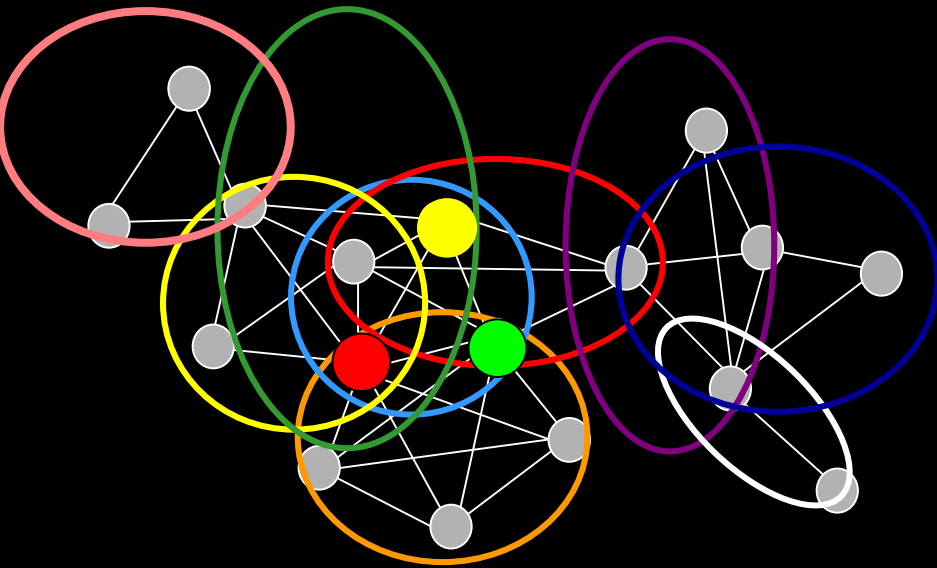


Clique tree

- Each tree node is a clique
- For every protein, the cliques that contain this protein form a connected subtree

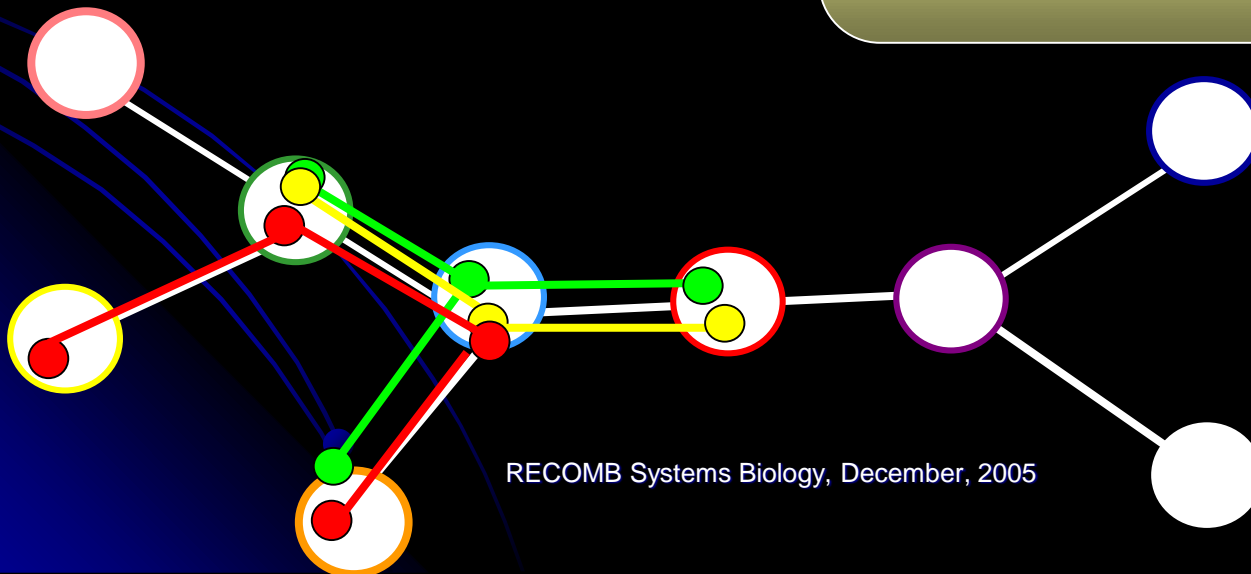


Key properties of a clique tree



We can trace each protein as it enters/ leaves each complex (protein group)

Can such a tree always be constructed?



Clique trees can be constructed only for **chordal graphs**

Chord = an edge connecting
two non-consecutive nodes of
a cycle

Chordal graph – every cycle of
length at least four has a chord.

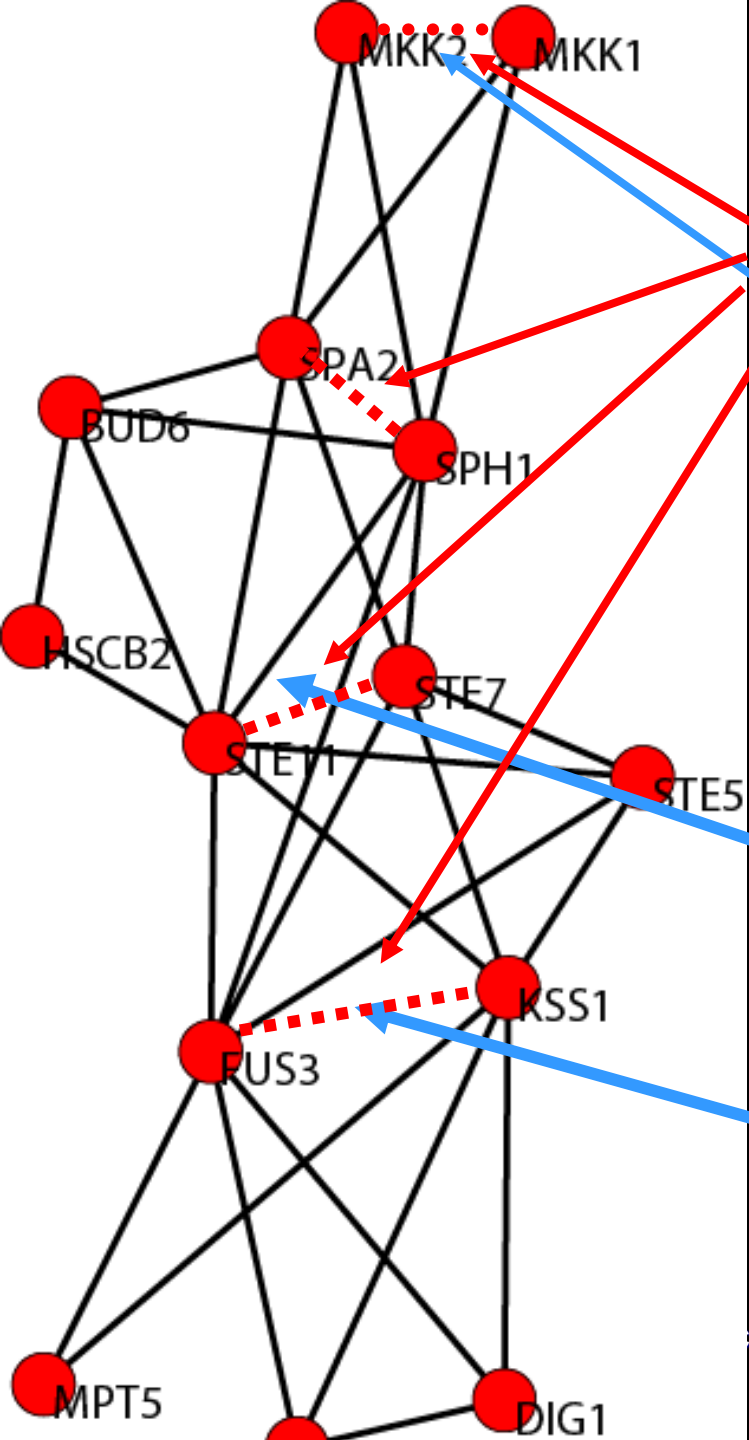
With these two edges the graph is
not chordal

hole

Chordal PPI-networks???

Do we ask for too much?

- If we take the whole PPI network this **IS asking too much.**
- If we take a densely connected subgraph identified as a “functional module” by any of a number of approaches **it is typically chordal** or close to it!



Add special “**OR**” edges

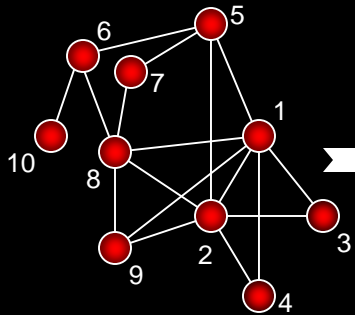
assembled by
Spirin et al. 2004

Square 1:
MKK1, MKK2 are
experimentally
confirmed to be **redundant**

Square 2:
STE11 and STE7 –
missing interaction

Square 3:
FUS3 and KSS1 –
similar roles (**replaceable**
but not redundant)

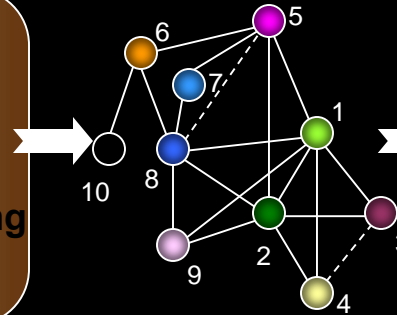
Original Graph, G



Graph modification

1. Add edges between nodes with identical set of neighbors
2. Eliminate *squares* (4-cycles) (if any) by adding a (restricted) set of “fill in” edges connecting nodes with similar set of neighbors

Modified Graph, G*



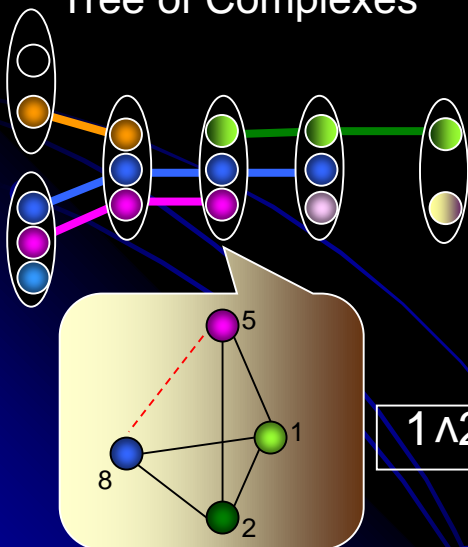
Is the modified graph chordal?

No

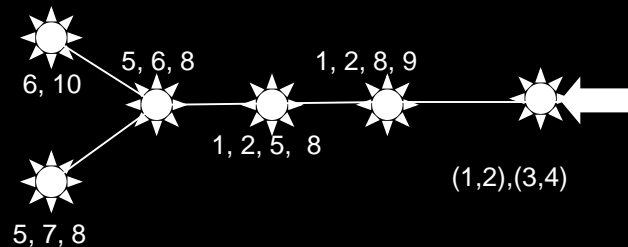
STOP

Yes

Tree of Complexes



Maximal Clique Tree of G*



1. Compute *perfect elimination order* (PEO)
2. Use PEO to find maximal cliques and compute *clique tree*

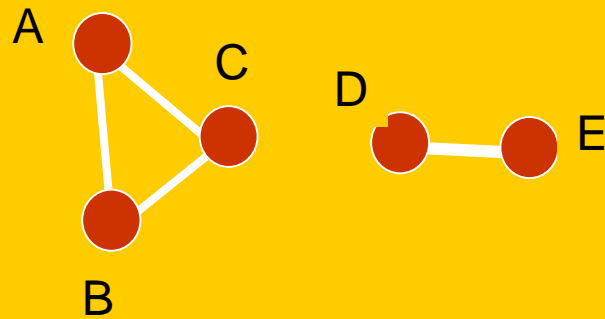
Protein

Fill-in edge

Maximal clique

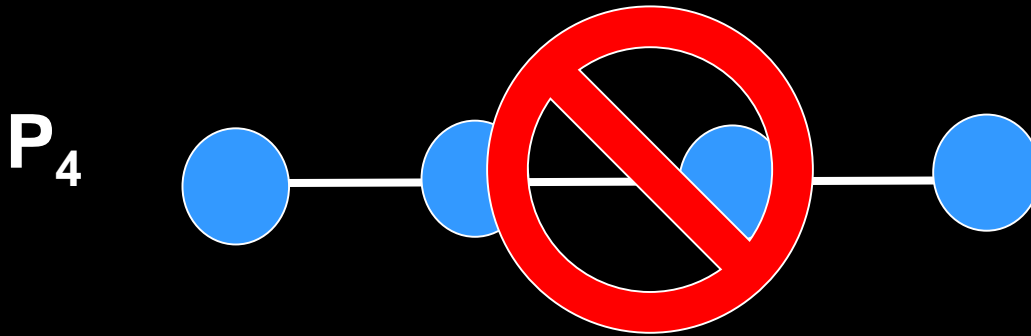
$1 \wedge 2 \wedge (5 \vee 8)$

Representing a functional group by a Boolean expression



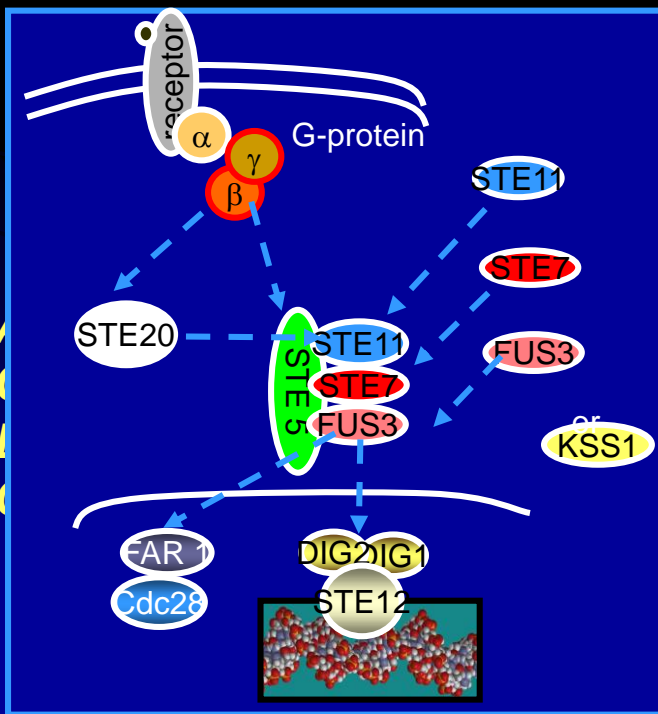
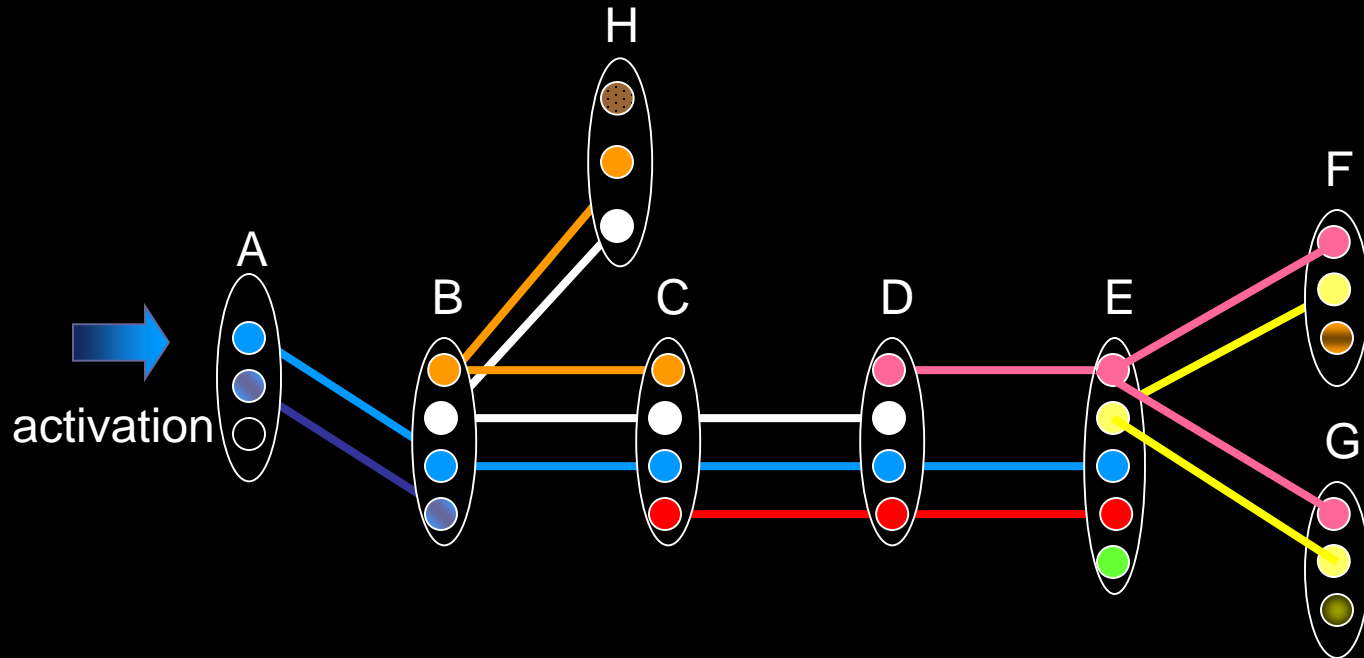
$$(A \wedge B \wedge C) \vee (D \wedge E)$$

Not all graphs can be represented by Boolean expression



Cographs = graphs which can
be
ex

Connected components of a cograph
are dense and “**cliquish**”



● = STE11

● = STE5

● = STE7

● = FUS3

● = KSS1

● = DIG1 \wedge DIG2

○ = HSCB2

● = BUD6

● = MPT5

FUNCTIONAL GROUPS

$B = BUD6 \wedge (SPH1 \vee SPA2) \wedge STE11$
 $D = SPH1 \wedge (STE11 \vee STE7) \wedge FUS3$
 $F = (FUS3 \vee KSS1) \wedge DIG1 \wedge DIG2$
 $H = (MKK1 \vee MKK2) \wedge (SPH1 \vee SPA2)$

Summary

- We proposed a new method delineating functional groups and representing their overlaps
- Each functional group is represented as a Boolean expression
- If functional groups represent dynamically changing protein associations, the method can suggest a possible order of these dynamic changes
- For static functional groups it provides compact tree representation of overlaps between such groups
- Can be used for predicting protein-protein interactions and putative associations and pathways
- To achieve our goal we used existing results from chordal graph theory and cograph theory but we also contributed new graph-theoretical results.

Future work

- So far we used methods developed by other groups to delineate functional modules and analyzed them or method. We are working on a new method which would work best with our technique.
- Considering possible ways of dealing with long cycles.
- Since fill-in process is not necessarily unique consider methods of exposing simultaneously possible variants.
- Add other information, e.g., co-expression in conjunction with our tree of complexes.

Acknowledgments

Elena Zotenko (UMD/NCBI)

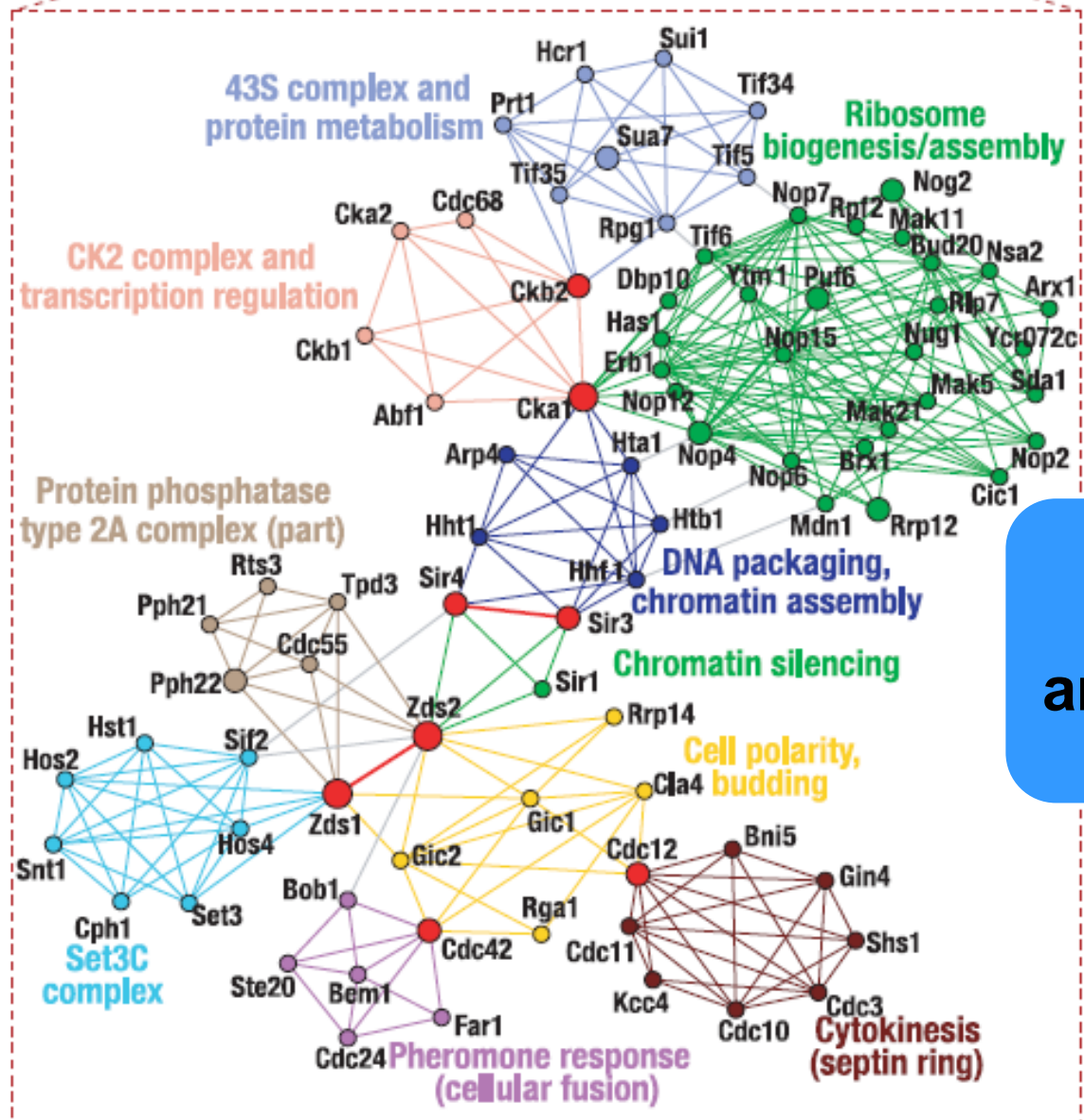
**Katia S. Guimaraes (Federal University
of Pernambuco, Brazil / NCBI)**

- **Raja Jothi (NCBI)**

**See also our
poster**

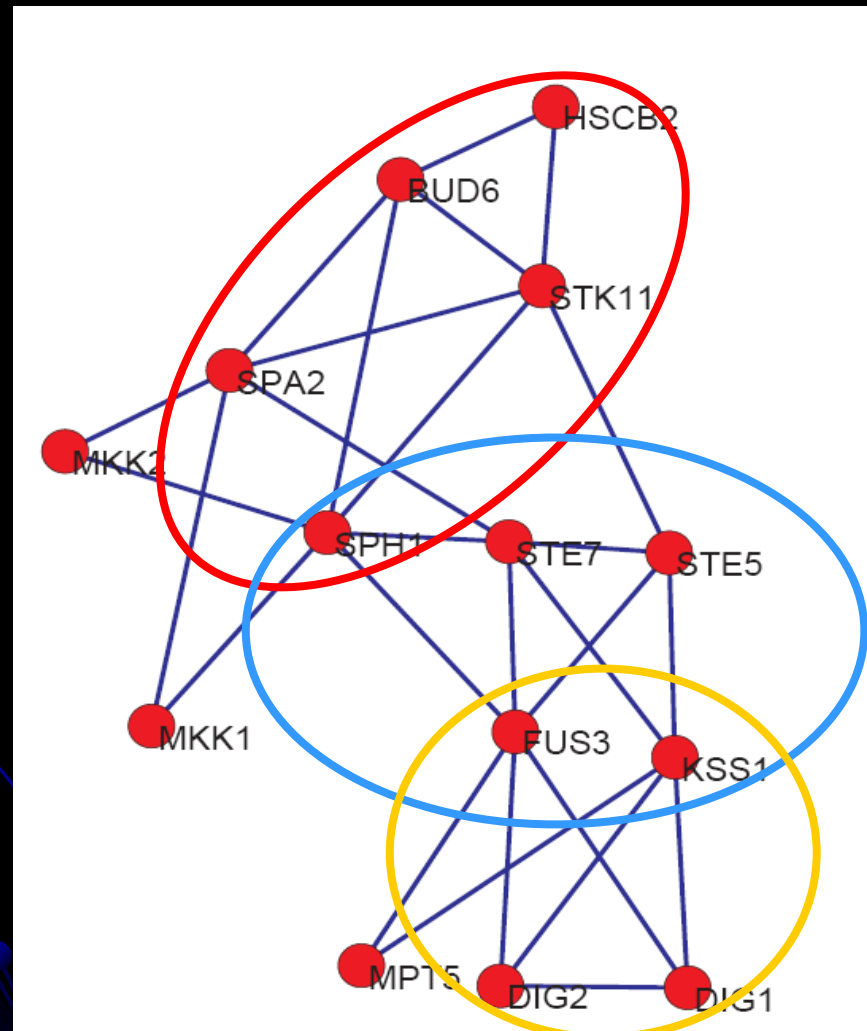
Applications

- “OR” edges – alternative/possible missing interactions. It is interesting to identify them and test which (if any) of the two possibilities holds
- Testing for consistency
- Hypothesis



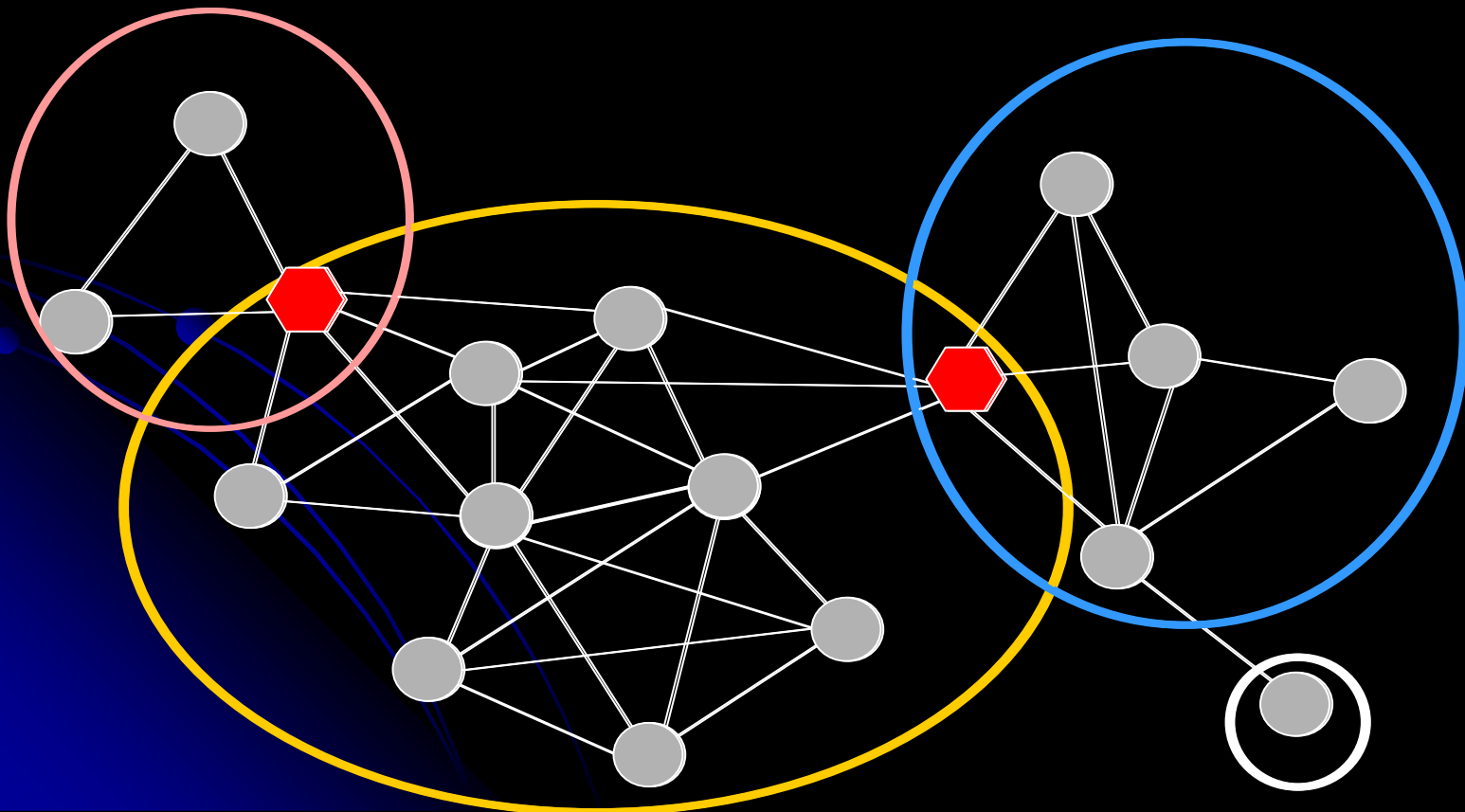
**Communities
are typically chordal !**

Community method applied to Pheromone data

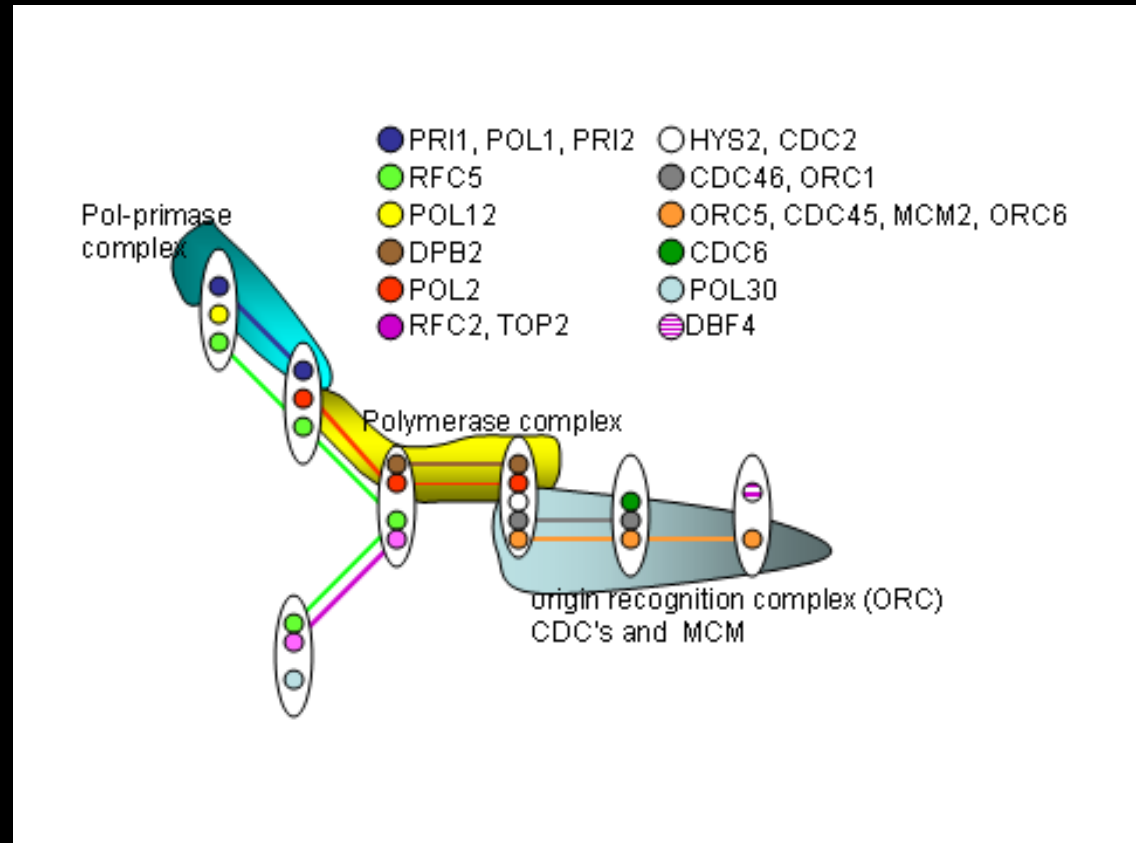
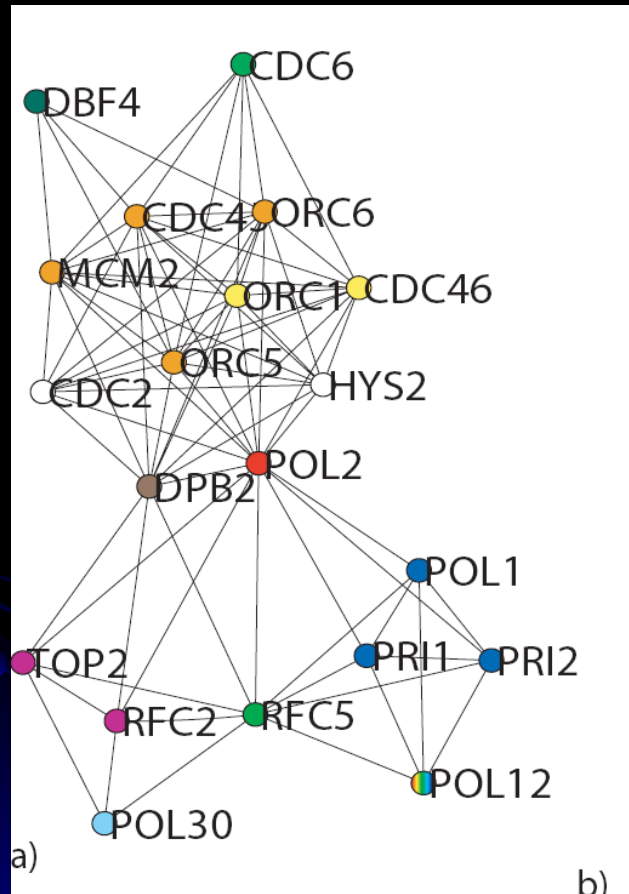


Palla et al.

- Identify functional modules (allow them to overlap)
 - For every node, record functional modules it belongs
- (Palla et al. Nature, June 2005)

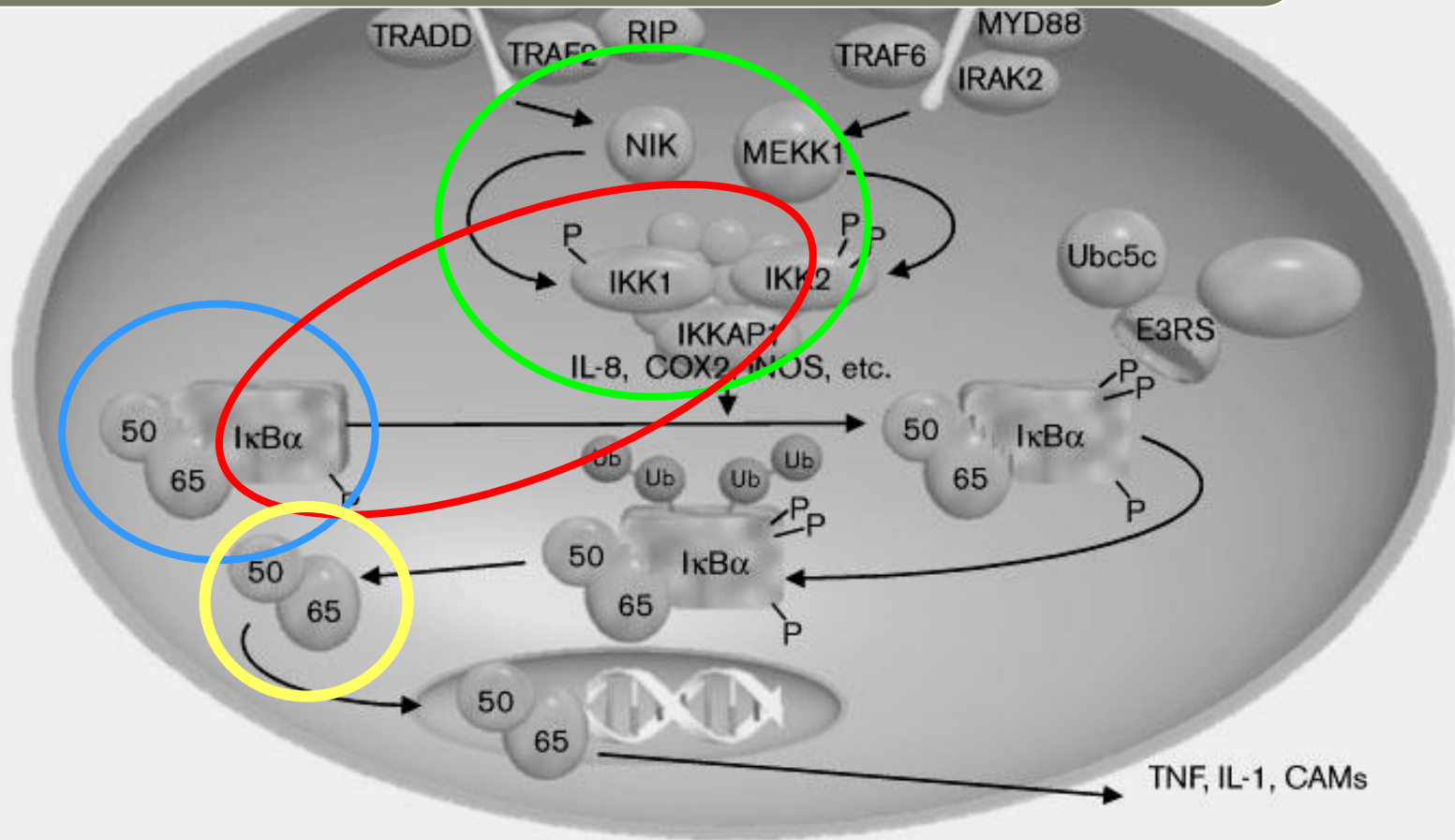


Transcription complex



NF- κ B resides in the cytosol bound to an inhibitor I κ B.
Binding of ligand to the receptor triggers signaling cascade
In particular phosphorylation of I κ B

I κ B then becomes ubiquitinated and destroyed by proteasomes.
This liberates NF- κ B so that it is now free to move into
the nucleus where it acts as a transcription factor



Based on network assembled by:
 Bouwmeester T, Bauch A, Ruffner H, Angrand PO, Bergamini G, Croughton K, Cruciat C, Eberhard D, Gagneur J, Ghidelli S, *et al.*:
 A physical and functional map of the human TNF-alpha/NF- κ B signal transduction pathway.
 (all paths of length at most 2 from NIK to NF- κ B are included)

